# Population Genetics of the Eurasian Bat (Myotis brandtii)

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Population structure defines relatedness between individuals that live in the same place or in different geographical locations. Individuals may be more related when they live in small groups or are isolated from other populations, due to inbreeding. Individuals from populations that are less related tend to be separated by long distances or geographical barriers such as mountains. An understanding of a species' population structure can help conserve species and understand how diseases spread between populations. This is particularly important in bats as they carry a lot of viruses that can infect humans, such as rabies. This study investigated population structure within the Eurasian Brandt's bat (*Myotis brandtii*) and subspecies found in Mongolia (*Myotis brandtii gracilis*). This study found a lack of population structure in the European populations but found the Mongolian population to be isolated from the others. Populations with increased distances between them were less related. However, the European populations still appear to be breeding with one another, suggesting a lack of geographical barriers. Male bats tend to travel further distances than female bats and how this affects population structure is an interesting focus for further study.

#### Abstract

Genetic studies of bat populations are important due to the wider implications within conservation and the control of transmissible diseases. Brandt's bats (*Myotis brandtii*) are widely distributed throughout Europe and Asia and exhibit swarming; a promiscuous mating system that reduces the risk of inbreeding. The aim of this study is to use seven microsatellite *loci*, polymorphic in *Myotis brandtii*, to determine the level of large-scale population structure across eight populations in Europe and Asia, including a population of the cryptic subspecies *Myotis brandtii gracilis*. The distance between sites is large and highly variable with multiple potential dispersal barriers. Genetic diversity between and within populations was measured using Hardy-Weinberg exact tests, F-statistics and the Mantel test assessed isolation by distance (IBD). The results of this study indicated a low level of population structure between the individuals sampled in Europe, with high levels of heterozygosity. The Mongolian population of suspected subspecies *M. b. gracilis*, was the most genetically differentiated from the other populations. Mantel tests found significant IBD between each population, with a gradual increase in genetic differentiation with geographical distance. Despite high IBD, evidence of high gene flow and heterozygosity suggests IBD is not considered a conservation issue in the Brandt's bat. This study stimulates the need for further investigation into sex-biased dispersal.

#### Introduction

Data obtained through population studies has implications within conservation biology. Knowledge of migration between sub-populations can help to predict the likely impacts of habitat destruction and segmentation. M. brandtii is currently listed as 'least concern' on the IUCN red list due to high abundance (1) and widespread distribution throughout Europe and Asia (2). However, in the event that a species becomes threatened or endangered, population studies can help make informed regarding breeding decisions programmes and reintroductions (3-4). Identifying the lack of population structure in the closely related Natterer's bats (Myotis nattereri) highlighted the importance of the identification and conservation of swarming sites, as they are invaluable in reducing inbreeding by increasing gene flow between sub-populations (5).

Population genetics also give an insight into movement patterns, aiding the development of accurate analytical models of the spread of disease (7). This is particularly important within bats as they are major natural reservoirs of several multi-host viruses (SARS-Like Coronaviruses, MERS-Like Coronaviruses, Nipah virus, Ebola virus, Rabies virus and Hendra viruses) (8). European Bat Lyssaviruses type 1 (EBLV-1), type 2 (EBLV-2) and Bokeloh Bat Lyssavirus (BBLV), are all causative agents of rabies in European bats, and have been isolated from several Myotis bat species (*M. daubentonii, M. dasycneme* and *M. nattereri*) (9). As a result of habitat disturbance, bats are living in closer proximity to humans and the infection of new host species is becoming more frequent, posing a higher risk for human infection (10). Studies also provide an insight into the epidemiology of diseases affecting bats. White-nose syndrome (WNS) is a fungal disease of North American bats and causes death in susceptible species (11). Population studies of little brown bats (*Myotis lucifugus*) suggested that population structure, derived from female migration patterns, influenced the spread of WNS across Pennsylvania and West Virginia (12).

In this study, seven microsatellite loci, polymorphic in M. brandtii (13-15) are analysed to identify genetic variation between and within sub-populations of *M. brandtii* and subspecies M. brandtii gracilis. This study will provide evidence to assess the level of population structure and isolation by distance in M. brandtii and discuss the implications. Ecological data of the Brandt's bat is limited due to cryptic morphology, nocturnality and ability of flight. M. brandtii morphologically resembles three other species (Myotis alcathoe, Myotis mystacinus and Myotis ikonnikovi) (16). M. brandtii is considered a small bat species with an adult body weight of 4-8 g and can live upwards of 40 years (2). They inhabit temperate broadleaf, mixed or coniferous woodland, typically near water (17), and feed on nonaquatic small insects and spiders (18). Summer roosts are being found more frequently near human habitation; in roofs and bird boxes, but more commonly in hollow trees. Hibernation locations tend to be in caves, cellars, mines and tunnels (19).

The Brandt's bat could show population substructure. A previous study found that Schreibers' long-fingered bat,



Figure 1. A small group of Brandt's bats roosting in a cave (A) and ventral photograph of a single Brandt's bat (B); reproduced from (55).

*Minipterus schreibersii natalensis*, demonstrated strong population substructure in the South, West and North-East regions of South Africa. Without obvious geographical barriers, the study concluded that the genetic diversity between the three sub-populations resulted from morphological differences and local biomes (20). However, due to the bats' flight ability, promiscuity and presence at swarming events, we predict *M.brandtii* will show little to no population structure, similar to that of Daubenton's bats (*Myotis daubentonii*) (21). Many studies have found a lack of population structure to be universal in seasonally migratory bats (22).



Figure 2. Red dots represent sampling locations across Europe. Map created for this study using mapping software [online] Available from: https://www.darrinward.com/lat-long/

#### Methods

A total of 135 bats were sampled from eight locations including England (Easegill Caverns), Germany (Mayener Grubenfeld), Switzerland (Gouffre de la Pleine Lune and Couffre Cathy within Parc Jurassien Vaudois), Finland (Turku), Latvia (Sikspārņu Cave), Russia (Petrozavodsk) and Mongolia (Ulan Bator) (Fig. 2). DNA was extracted and purified from 3 mm wing punches (stored in 70% ethanol at -80°C) using the Qiagen DNeasy Blood and Tissue Kit. Wing punches (3 mm) do not disrupt flight and heal completely in 2-3 weeks (23). The yield of genomic DNA was measured using the Thermo Fisher Invitrogen Qubit Fluorometer. Single template multiplex PCR was carried out with a final volume of 10  $\mu$ l, containing: Qiagen Multiplex PCR Master Mix, forward and reverse primers (fluorescently labelled) (Table 1), Genomic DNA, and

dH<sub>2</sub>O. Thermal cycling programme: 95 °C for 5 minutes followed by 28 cycles (95 °C for 30 seconds, 59 °C for 90 seconds and 72 °C for 30 seconds and lastly, 60 °C for 30 minutes). The PCR products were diluted with water (1:100) and amplicons were separated by capillary electrophoresis on the Applied Biosciences 3500xL Genetic Analyzer. Genemapper V software (24) was used to size and assign alleles.

For each sample site, observed heterozygosity (Ho), expected heterozygosity (He), allelic richness and effective number of alleles (Ae) was calculated using GENALEX 6.5 (25) add-in for Microsoft Excel. Guo and Thompson's (26) exact Hardy-Weinberg test was used to assess compliance with Hardy-Weinberg Equilibrium (HWE) for each locus in each population using the population genetics program GENEPOP 4.6 (27). Hardy-Weinberg exact tests are performed frequently in population genetics as deviations from HWE may highlight problems such as selection bias (28) and genotyping errors (29). The Markov Chain Method is used to estimate the exact P-values for each locus in individual populations and Fisher's method was used to calculate the overall P-value of all loci in each population. Pairwise FST values were calculated using GENEPOP 4.6 (27). F<sub>ST</sub> (the fixation index) is the value for genetic distance. It is proportional to the level of inbreeding within subpopulations relative to the metapopulation (30). The inbreeding coefficient (FIS) of an individual relative to the subpopulation were calculated using GENALEX using the Weir & Cockerham (31) method. A negative FIS value suggests individuals are less related to each other and a positive FIS value suggests individuals are less related. Isolation by distance is assessed on GENEALEX using the Mantel test, which compares genetic distance and geographical distance. Statistical significance of R (mantel coefficient) was attained by forming 999 permutations.

Ethical approval for this study was obtained by my supervisor, Professor Steve Paterson.

#### Results

The yield of genomic DNA, extracted from 3 mm wing punches, ranged from  $1.9-220 \text{ ng/}\mu\text{l}$ , with a mean final concentration of  $20.2 \text{ ng/}\mu\text{l}$ .

### Research

Locus	Sequences (5'-3') Forward (F) and Reverse (R) Primer	Array	Fluorescent Tag	Allele Size Range (bp)	Marker concentration (mM)
A24-Mluc (13)	F: GTGGTATGAAATAACCAGTTCACTTTG R: GTTTCAGACTGCATTACTGAAGAAATTATGG	(AC)n	FAM	473-491	0.2
Clone A2-Mluc	F: TGGCCCATGCTCATCATC R: GTTTCTGGTCTCAACTGGGTGCTC	(CA)n	VIC	91-135	0.05
D9 <sup>(14)</sup>	F: GTTTCTTTCCTCCCCTGTGCTC R: TCTGGACCCAAAATGCAGG	(CT)n	NED	120-150	0.2
E24 <sup>(14)</sup>	F: GTTTGCAGGTTCAATCCCTGACC R: AAAGCCAGACTCCAAATTCTG	(TC)n	FAM	215-253	0.2
ES43-Mluc <sup>(13)</sup>	F:GTTTAAGGGGGAGAGGAGTGG R:GCTGCGTGTCCAGAGG	(AC)n	FAM	377-403	0.2
H29 <sup>(15)</sup>	F: GTTTCAGGTGAGGATTGAAAACAC R: GCTTTATTTAGCATTGGAGAGC	(CA)n	FAM	170-202	0.4
Paur 6 <sup>(16)</sup>	F: GATCAGATTTCCAAACAGAG R: GTTTAGGTTCTTTCTTCAGCTATG	(AC)n(AG)n	PET	156-186	0.2

Expected heterozygosity

0 703

0.694

0.641

0.661

0.652

0.670

0.803

Table 1. Adapted from (22). Forward and reverse primer sequences, arrays, fluorescent tag, allele size range (bp) and marker concentration (mM) for the microsatellite *loci* used in this study. Allele sizes adapted for *M.brandtii*.

Table 2. Genetic diversity indices averaged across microsatellite *loci* for each population and the HWE *P*-values for exact tests. n= sample size.

Table 3. HWE p values for each locus in each individual population calculated using Markov chain method. The significant values are underlined (P < 0.01) and in boldface (P < 0.05).

Sample Site A24-Mluc Clone A2-Mluc D9 E24 ES43-Mluc H29 Paur6 England 0 4286 0.3708 0.9683 0 2280 0 0226 1 0000 0 6281 Germany 0.0388 0.1466 0.1358 0.0656 0.4036 0.6770 0.5965 0.5689 0.0945 0.5664 0.5725 0.6693 Switzerland 0.0386 0.1211 Finland 0.0035 0.0608 0.1792 0.0116 0.7078 0.0005 Latvia 1.0000 0.1308 0.8831 0.6416 0.0051 1.0000 0.5691 Russia 0 5046 0.0060 0 8908 0.3585 0.7923 1.0000 0 9758 0.8804 Mongolia 0.1413 0.8094 0.7102 0.0000 0.04041 0.8657

### **Population Diversity**

Sample site

England

Germany

Switzerlan

Finland

Latvia

Russia

Mongolia

The observed heterozygosity in each population ranged from 0.584-0.842 (Table 2). One population (Finnish) out of six deviated significantly from HWE for P<0.05. Three loci deviated significantly from HWE in the Finland population (P<0.05) as opposed to a maximum of two loci in other populations (Table 3). Russia and Mongolia had higher observed heterozygosity (0.702 and 0.842, respectively) than expected (0.670 and 0.803) whilst the other populations had lower observed heterozygosity than expected (Table 3). Mongolia and Russia also had the least number of positive FIS values (two loci) inferring higher levels of inbreeding than expected as opposed to a minimum of three loci in the other populations. Positive inbreeding coefficients (FIS) were found in all populations at a maximum of six loci (Finland). The highest inbreeding coefficient (1.000) was found at the Finnish site for the locus A24-Mluc (Table 4), where all individuals are homozygous. The lowest inbreeding coefficient (-0.3333) was found at the Russian site (Table 4).

Observed heterozygosity

0.649

0.686

0.621

0.584

0.634

0.702

0.842

n

21

33

23

20

13

#### **Population Differentiation**

Pairwise genetic distance ( $F_{ST}$ ) and geographical distance are presented in Table 5.  $F_{ST}$  values range from 0.0055 (Latvia and Germany) to 0.1662 (Finland and Mongolia). The largest linear geographical distance (7018 km) exists between Switzerland and Mongolia. The Mantel test found a positive correlation between genetic distance and geographical distance (Mantel test: R= 0.416, P< 0.001) showing significant isolation by distance.

HWE p

0.3638

0.0540

0.2198

< 0.001

0.2613

0.4213

0.9192

4 67

5.05

4.73

4.42

4.96

4.41

5.96

#### Discussion

Allelic Richness

7.14

10.00

7.71

9.14

8.57

6.29

10.00

#### **Population Diversity**

Each population had high heterozygosity across loci. One of the possible explanations for high individual population heterozygosity values in *M. brandtii* is that they are one of many bat species present during swarming (32-34). Swarming is a phenomenon that reduces the risk of inbreeding, taking place in late Summer and early Autumn, where large numbers of bats visit hibernacula (34). Currently, there is no consensus as to why bats swarm but there are multiple hypotheses. Three principal theories include opportunities to mate (35), information transfer from parental bats to offspring and, to assess suitable hibernacula (34). Opportunity to mate as a hypothesis has the most supporting evidence. Bats display significant social vocalisation, chasing and copulatory behaviour during this period (36). There is significant evidence to suggest swarming events increase gene flow between subpopulations, increasing genetic diversity in progeny (32).

Bats also demonstrate reproductive strategies known to increase genetic diversity within other animal species. The behaviours demonstrated by *M. brandtii* during mating periods are described as indiscriminate and promiscuous

Sample Site	A24- Mluc	Clone A2-Mluc	D9	E24	ES43-Mluc	H29	Paur6
England	0.5714	0.1765	-0.0026	0.2139	0. <b>1504</b>	-0.1799	-0.0696
Germany	0.1754	0.1384	-0.0999	0.1537	-0.0613	-0.0604	0.0000
Switzerland	-	0.2636	-0.0649	0.0686	0.1304	0.1493	-0.1020
Finland	1.0000	0.3070	-0.0360	0.1273	0.0523	0.2222	0.0213
Latvia	-0.1053	0.0788	-0.1362	0.0917	0.1268	-0.1347	0.0710
Russia	-0.3333	0.2809	-0.0602	0.0000	-0.0614	-0.0588	-0.0254
Mongolia	-0.0162	-0.0859	-0.1027	0.0403	0.0949	-0.0897	-0.0335

Finland

1628

1465

1873

0.0083

0.0198

0 1662

Latvia

1764

1413

1773

391.6

0.0100

0.1590

Russia

2292

2085

2468

665.5

709.4

0.1554

Mongolia

6880

6678

7018

5276

5265

4616

Switzerland

1057

436.4

0.0161

0.0139

0.0212

0.1645

**Table 4.** F<sub>IS</sub> values at each *loci* for each population. Results that infer higher levels of inbreeding than expected are in boldface (31).

Table 5. Pairwise  $F_{ST}$ values and distance between populations (km). Distances (km) are above the diagonal.  $F_{ST}$ values are below the diagonal.

(35). After copulation, females store spermatozoa within their reproductive tract, delaying ovulation and fertilisation until Spring (35, 37). Studies on dissimilar species have demonstrated that sperm storage and promiscuity increase genetic diversity, for example, in the *Anolis sagrei* lizard (38) and passerine birds (39).

Germany

790.3

0.0204

0.0153

0.0055

0.0200

0.1622

Despite populations showing high observed heterozygosity and not deviating significantly from HWE, each population had positive inbreeding coefficients ( $F_{IS}$ ) at various *loci*, particularly in three *loci* (Clone A2-Mluc, E24 and ES43 Mluc). This contradiction could be attributed to sampling bias, small sample sizes and null alelles (40).

### **Population Differentiation**

Sample Site

England

Germany

Switzerland

Finland

Latvia

Russia

Mongolia

England

0.0214

0.0272

0.0195

0.0123

0 0241

0.1488

Pairwise European  $F_{ST}$  values indicated little genetic differentiation between populations, using guidelines for interpretation (41), indicating low population structure in the Brandt's bat. The results suggested there are no significant barriers to gene flow in Europe. These findings are in agreement with other population genetic studies on seasonally migratory bats such as the Daubenton's bat (21) and wide ranging species such as the Regent Honeyeater bird, *Anthochaera phrygia* (42). The Mongolian population had high pairwise  $F_{ST}$  values with all other populations, suggesting they are isolated from the others and not interbreeding.

Low  $F_{ST}$  values suggest populations are not significantly genetically distinct. However significant IBD presents a barrier to gene flow. As distance increases, populations become more genetically diverse. Geographical barriers of movement in the Brandt's bats may include mountain ranges and large expanses of different climates (deserts). Bright lights and disruption of vision can affect bat migration and tracking long-range movements (43) suggesting brightly lit cities and roads have the potential to restrict geneflow. However, migratory behaviour which can reach up to 618 km (43-45), and flight abilities may be the reason for the apparent lack of barriers to gene flow between sub-populations.

Similarly to the apparent lack of geographic barriers found in this study, Castella et al. (14) also identified a lack of population structure between two populations of Myotis myotis on either side of the Gibralter Strait (14 km wide stretch of water separating peninsular Spain and Gibralter from Morocco and Ceuta in Africa). However, further analysis of a mitochondrial gene (cytochrome b) confirmed the two populations were genetically distinct with no interbreeding over the Strait of Gibralter. This suggests other methods of molecular analysis on the European populations of *M. brandtii* are required to definitively state whether they are interbreeding. Myotis bats also demonstrate a lack of site fidelity, changing roosts frequently depending on their reproductive status (35, 46), encouraging interbreeding between sub-populations. Male bats are recorded to disperse further than females. Females tend to be more philopatric as they have to exploit resources and provide parental care to immobile progeny (21, 47). Population structures generally fall into three categories - populations divided by age, sex or social/geographic factors (48). This study analysed geographic factors suggesting further investigation into age and gender diversity indices is necessary to determine sex barriers on population structure.

# Implications within conservation and Disease transmission

Low genetic distance between populations suggests a lack of geographical barriers within Europe which is promising for bat conservation. This study highlights the importance of protecting swarming sites as swarming species have higher gene flow (49). A major feature currently concerning the conservation of many species is climate change. Studies have indicated that European bats may face a serious threat with climate change (50) and may respond with a population shift northwards, breeding earlier (51) or changing migration times (52). This study confirms the widespread distribution of *M. brandtii* in temperate regions and tropical climates (Mongolia) suggesting they are able to adapt to changing climates, providing there is available habitat northwards. Migratory bats are known to migrate over open sea as they are frequently found on remote islands, ships and oil rigs (53). This study indicates that British bat populations are interbreeding with continental European populations; presenting an interesting insight into the spread of diseases, such as the rabies virus. Many studies focus on the control of dogs and foxes to control the spread of rabies, and highlight rabies-free status in some European countries. However, rabies is a trans-boundary disease due to the migratory nature of bats, and the importation of infected animals and reinfections have occurred in Italy, Greece and Slovakia (54). This highlights the need for trans-boundary and species-wide approaches to controlling the spread of rabies.

#### Conclusions

In summary, European populations of M. brandtii show low population structure but significant isolation by distance. The high F<sub>ST</sub> values between the Mongolian population of M. b. gracilis suggest they are isolated and not breeding with the other populations. Similar to the Gibralter Strait study (14), further molecular analyses could confirm or refute the lack of population structure in Europe. There are a number of possible reasons for high heterozygosity and low FST values in the European populations, despite significant isolation by distance, such as: a lack of physical boundaries, ability of flight, migratory behaviour, swarming, lack of site fidelity, sex-biased dispersal and promiscuity. Similar literature suggests a study on sex-biased dispersal would provide further insight into the population structure of Brandt's bats. These findings highlight the importance of conserving swarming sites to reduce inbreeding, and the possible consequences to climate change. They also support the idea of trans-boundary disease control strategies due to the migratory nature of bats.

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#### References

- Hutson AM, Spitzenberger F, Coroiu I, Aulagnier S, Juste J, Karatas A, et al. Myotis brandtii. 2008. http:// www.iucnredlist.org/details/ full/14125/0 (accessed 8 Oct 2017).
- Seim I, Fang X, Xiong Z, Lobanov AV, Huang Z, Ma S, et al. Genome analysis reveals insights into physiology and longevity of the Brandt's bat *Myotis brandtii*. Nat Commun 2013;4(2212). Doi: 10.1038/ ncomms3212.
- Avise JC. Molecular population structure and the biogeographic history of a regional fauna: A case history with lessons for conservation biology. *Oikos* 1992;63(1):62-76.
- Manel S, Schwartz MK, Luikart G and Taberlet P. Landscape genetics: Combining landscape ecology and population genetics. *Trends Ecol Evol* 2003;18(4):189-197.
- Rivers NM, Butlin RK and Altringham JD. Genetic population structure of natterer's bats explained by mating at swarming sites and philopatry. *Mol Ecol* 2005;14(14):4299-312.
- 6. Bowen BW, Bass AL, Soares L and Toonen RJ. Conservation

implications of complex population structure: lessons from the loggerhead turtle (*Caretta caretta*). *Mol Ecol* 2005;14(8):2399-402.

- Sattenspiel L. Population structure and the spread of disease. *Hum Biol* 1987;59(3):411-38.
- Han H, Wen HL, Zhou CM, Chen FF, Luo LM, Liu JW, *et al.* Bats as reservoirs of severe emerging infectious diseases. *Virus Res* 2015;205:1-6.
- Schatz J, Fooks A, McElhinney L, Horton D, Echevarria J, Vazquez-Moron S, *et al.* Bat rabies surveillance in Europe. *Zoonoses Public Health* 2013;60(1):22-34.
- Wibbelt G, Moore MS, Schountz T and Voigt CC. Emerging diseases in Chiroptera: Why bats? *Biol Lett* 2010;6(4):438-40.
- Warnecke L, Turner JM, Bollinger TB, Misra V, Cryan PM, Blehert DS, et al. Pathophysiology of white-nose syndrome in bats: A mechanistic model linking wing damage to mortality. *Biol Lett* 2013;9(4):20130177.
- Miller-Butterworth CM, Vonhof MJ, Rosenstern J, Turner GG and Russel AL. Genetic structure of littler brown bats (*Myotis lucifugus*) corresponds with spread of white-nose syndrome among hibernacula. *J Hered* 2014;105(3):354-364.
- Jan C, Dawson DA, Altringham JD, Burke T and Butlin RK. Development of conserved microsatellite markers of high crossspecies utility in bat species (Vespertilionidae, Chiroptera, Mammalia). *Mol Ecol Resour* 2012;12(3):532-48.
- Castella V, Ruedi M, Excoffier L and Ibanez C. Is the Gibralter strait a barrier to gene flow for the bat *Myotis myotis* (Chiroptera: Vespertilionidae). *Mol Ecol* 2000;9(11):1761-72.
- Burland TM, Barratt EM and Racey PA. Isolation and characterization of microsatellite loci in the brown long-eared bat, *Plecotus auritus*, and cross-species amplification within the family Vespertilionidae. *Mol Ecol* 1998;7(1):136-8.
- von Helverson O, Heller K-G, Mayer F and Nemeth A. Cryptic mammalian species: a new species of whiskered bat (*Myotis alcathoe* n. sp.) in Europe. *Naturwissenschaften* 2001;88(5):213-23.
- Tsytsulina K. Myotis ikonnikovi (Chiroptera, Vespertilionidae) and its relationships with similar species. Acta Chiropterol 2001;3(1):11-19.
- Natural England. Focus on Bats discovering their lifestyle and habitats (NE23). 2007. https:// webarchive.nationalarchives.gov.uk/20150303022058/http:// publications.naturalengland.org.uk/file/155449 (accessed 18 Oct 2017).
- 19. Schober W, Grimmberger E. *A guide to bats of Britain and Europe*. London: Hamlyn Publishing Group Limited, 1987.
- Miller-Butterworth CM, Jacobs DS and Harley EH. Strong population substructure is correlated with morphology and ecology in a migratory bat. *Nature* 2003;424(6945);187-91.
- Wright S. Evolution and the Genetics of Populations. Variability Within and Among Natural Populations. Vol. 4. Chicago: University of Chicago Press, 1978.
- Laine VN, Lilley TM, Norrdahl K and Primmer CR. Population genetics of Daubenton's bat (*Myotis daubentonii*) in the Archipelago Sea, SW Finland. ANN ZOOL FENN 2013; 50(5):303-315.
- Burland TM and Wilmer JW. Seeing in the dark: molecular approaches to the study of bat populations. *Biol Rev* 2001; 76(3):389-409.
- Weaver KN, Alfano SE, Kronquist AR and Reeder DM. Healing rates of wing punch wounds in free-ranging little brown Myotis (*Myotis lucifucgus*). Acta Chiropterol 2009; 11(1):220-223.
- 25. Applied Biosystems. 2012. Genemapper Software 5. Foster City, CA.
- Peakall R and Smouse PE. GenAlEx 6.5: genetic analysis in Excel. Population genetic software for teaching and research. *Bioinformatics* 2012; 28(19):2537-2539.
- Guo SW and Thompson EA. Performing the exact test of Hardy-Weinberg proportion for multiple alleles. *Biometrics* 1992; 48(2):361-372.
- Rousset F. Genepop'007: A complete reimplementation of the Genepop software for Windows and Linux. *Mol Ecol Resour* 2008; 8(1):103-106.
- Schaid DJ and Jacobsen SJ. Biased tests of association: comparisons of allele frequencies when departing from Hardy-Weinberg proportions. Am J Epidemiol 1999; 149(8):706-711.
- Xu J, Turner A, Little J, Bleecker ER and Meyers DA. Positive results in association studies are associated with departure from Hardy-Weinberg equilibrium: hint for genotyping error?. *Hum Genet* 2002; 111(6):573-573.
- Weir BS and Cockerham CC. Estimating F-statistics for the analysis of population structure. *Evolution* 1984; 38(6):1358-1370.
- 32. Beebee T and Rowe G. *An Introduction to Molecular Ecology*. 2nd ed. New York: Oxford University Press, 2008.
- Parsons KN, Jones G, Davidson-Watts I and Greenaway F. Swarming of bats at underground sites in Britain – implications for conservation.

Biol Conserv 2003; 111(1):63-70.

- Vlaschenko A, Hukov V, Naglov A, Prylutska A, Kravchenko K and Rodenko O. Contribution to ecology of Brandt's bat, *Myotis brandtii* (Chiroptera, Vespertilionidae), in the North -Eastern Ukraine: comparison of local summer and winter bat assemblages. *Vestn* zoologii 2016; 50(3):231-240.
- Piksa K. Swarming of *Myotis mystacinus* and other bat species at high elevation in the Tatra Mountains, Southern Poland. *Acta Chiropterol* 2008; 10(1):69-79.
- Thomas DW, Fenton F and Barclay RMR. Social behaviour of the little brown bat, *Myotis lucifugus*: 1. Mating Behaviour. *Behav Eco and Soc bio* 1979; 6(2):129-136.
- Bogdanowicz W, Piksa K and Tereba A. Genetic structure in three species of whiskered bats (*Genus Myotis*) during swarming. J Mammal 2012; 93(3):799-807.
- Pfeiffer B and Mayer F. Spermatogenesis, sperm storage and reproductive timing in bats. *J Zool* 2013; 289(2):77-85.
- Calsbeek R, Bonneaud C, Prabhu S, Manoukis N and Smith TB. Multiple paternity and sperm storage lead to increased genetic diversity in Anolis lizards. *Evol Ecol Res* 2007; 9(3):495-503.
- Gohli J, Anmarkrud JA, Johnsen A, Kleven O, Borge T and Lifjeld JT. Female promiscuity is positively associated with neutral and selected genetic diversity in passerine birds. *Evolution* 2013; 67(5):1406-1419.
- 41. Chapuis M and Estoup A. Microsatellite Null Alleles and Estimation of Population Differentiation. *Mol Biol Evol* 2007; 24 (3):621-631.
- Kvistad L, Ingwersen D, Pavlova A, Bull JK and Sunnucks P. Very low population structure in a highly mobile and wide-ranging endangered bird species. *PloS one* 2015;10(12).
- 43. Mathews F, Roche N, Aughney T, Jones N, Day J, Baker J and Langton S. Barriers and benefits: implications of artificial night-lighting for the distribution of common bats in Britain and Ireland. *Philos T Roy Soc B* 2015; 370(1667):20140124.
- Hutterer R, Ivanova T, Meyer-Cords C and Rodrigues L. Bat migrations in Europe. A review of banding data and literature. Bonn: Federal Agency for Nature Conservation, 2005. [Out of print]

- Popa-Lisseanu AG and Voigt CC. Bats on the move. *J Mammal* 2009; 90(6):1283- 1289.
- Ngamprasertwong T, Piertney SB, Mackie I and Racey PA. Roosting habits of Daubenton's bat (*Myotis daubentonii*) during reproduction differs between adjacent river valleys. *Acta Chiropterol* 2014; 16 (2):337-47.
- Wolff J. What is the role of adults in mammalian juvenile dispersal? Oikos 1993; 68:173-176.
- Sattenspiel L and Simon CP. The spread and persistence of infectious diseases in structured populations. *Math Bio Sci* 1988; 90 (1-2):341-366.
- Burns LE, Frasier TR and Broders HG. Genetic connectivity among swarming sited in the wide ranging and recently declining little brown bat (*Myotis lucifugus*). Ecol Evol 2014; 4(21):4130-4149.
- Rebelo H, Tarroso P and Jones G. Predicted impact of climate change on European bats in relation to their biogeographic patterns. *Glob Change Biol* 2010: 16(2):561-576.
- 51. Parmesan C. Ecological and evolutionary responses to recent climate change. *Annu Rev Ecol Evol S* 2006; 37:637-669.
- Miller-Rushing AJ, Lloyd-Evans TL, Primack RB and Satzinger P. Bird migration times, climate change, and changing population sizes. *Glob Change Biol* 2008; 14:1959-1972.
- Ahlén I, Baagøe LB and Bach L. Behavior of Scandinavian bats during migration and foraging at sea. J Mammal 2009; 90(6):1318 –1323.
- Cliquet F, Freuling C, Smreczak M, Van der Poel WH, Horton D, Fooks AR, et al. Development of harmonised schemes for monitoring and reporting of rabies in animals in the European Union. EFSA Supporting Publications 2010;7(7):67E.
- Podlutsky AJ, Khritankov AM, Ovodov ND and Austad SN. A new field record for bat longevity. *The J Gerontol A Biol Sci Med Sci* 2005; 60 (11):1366-68

## Driving is a risky business!



Dr Laura Bonnett (2020 William Guy Lecturer)

Wednesday 2nd December 2020 15.30-17.00



Leggate Theatre, Victoria Gallery and Museum (Building 421 in grid square D7 on campus map)

Named in honour of William Augustus Guy, an early medical statistician and past RSS president, this prestigious volunteer role recognises fellows with a successful track record in undertaking school outreach activities. Over 2020, Laura (pictured) will be delivering lectures to statisticians and students across the UK on the topic of 'Driving is a risky business!' which covers various aspects of road safety where statistics can illuminate people's decisions. The lecture will include a discussion regarding conditional probabilities, together with ethical considerations, which will appeal across the curricula.

For talk abstract, and registration details please go to the RSS Merseyside website https://sites.google.com/site/rssmerseyside/research-meetings/william-guy-lecturer

